

# T4 DNA Polymerase (5,000 U/ml)



**Catalog:** RK20539

**Size:** 200 U / 1000 U

**Concentration:** 5,000 U/ml

**Components:**

|                                |         |
|--------------------------------|---------|
| T4 DNA Polymerase (5,000 U/ml) | RM21302 |
| 10X ABuffer B                  | RM20126 |

## Product Description

T4 DNA Polymerase catalyzes the synthesis of DNA in the 5' → 3' direction and requires the presence of template and primer. This enzyme has a 3' → 5' exonuclease activity which is much more active than that found in DNA Polymerase I (*E.coli*). Unlike *E.coli* DNA Polymerase I, T4 DNA Polymerase does not have a 5' → 3' exonuclease function.

It is applicable to 3' overhang removal to form blunt ends, 5' overhang fill-in to form blunt ends, single strand deletion subcloning, second strand synthesis in site-directed mutagenesis and probe labeling using replacement synthesis.

**Product Source:** Purified from a strain of *E.coli* that carries the T4 DNA Polymerase gene.

**Unit Definition:** One unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTP into acid insoluble material in 30 minutes at 37°C.

**Storage Conditions:** 100 mM KPO<sub>4</sub>, 1 mM DTT, 50% Glycerol, pH 6.5 @ 25°C

**Storage Temperature:** -20°C

**Reaction Conditions:** 1X ABuffer B, Incubate at 12°C.

**1X ABuffer B:** 10 mM Tris-HCl, 50 mM NaCl, 10 mM MgCl<sub>2</sub>, 1 mM DTT, pH7.9 @ 25°C

**Activity in ABclonal Buffer**

| ABufferA | ABufferB | ABufferC | ABufferS |
|----------|----------|----------|----------|
| 60%      | 100%     | 100%     | 100%     |

**Heat Inactivation:** 75°C for 20 min

**Molecular Weight:** Theoretical 104000 daltons

**5' - 3' Exonuclease:** No

**3' - 5' Exonuclease:** Yes

**Strand Displacement:** No

**Error Rate:** ~ 1 x 10<sup>-6</sup> bases

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## Instructions

### Protocol for blunting ends by 3' overhang removal and 3' recessed (5' overhang) end fill-in using T4 DNA Polymerase:

1. DNA should be dissolved in 1X reaction buffer\* supplemented with 100 μM dNTPs.
2. Add 1 unit T4 DNA Polymerase per microgram DNA.
3. Incubate 15 minutes at 12°C.
4. Stop reaction by adding EDTA to a final concentration of 10 mM and heating to 75°C for 20 minutes.

**CAUTION:** Elevated temperatures, excessive amounts of enzyme, failure to supplement with dNTPs or long reaction times will result in recessed ends due to the 3' → 5' exonuclease activity of the enzyme.

*\*T4 DNA Polymerase can be used in CutA, CutB and CutS as well as ABuffer A/B/S and T4 DNA Ligase Reaction Buffer. Optimal activity is observed in ABufferB. BSA supplementation is recommended when using a buffer that does not already contain BSA.*

### Notes:

1. For fill-in reactions only: T4 DNA Polymerase can be used in CutA, CutB and CutS as well as ABuffer A/B/S and T4 DNA Ligase Reaction Buffer.
2. For blunting reactions requiring removal of overhangs: T4 DNA Polymerase can be used in CutA, CutB and CutS as well as ABuffer A/B/S and T4 DNA Ligase Reaction Buffer. CutC and ABuffer C are not recommended when overhang removal is required.
3. Optimal activity is observed in ABuffer B.
4. Supplement with dNTPs\*.
5. BSA supplementation is recommended when using a buffer that does not already contain BSA.
6. Incubate at temperature suggested for specific protocol.

*\* Refer to specific protocol to determine recommended dNTP concentrations.*

### QC Process:

- Purity (SDS-PAGE) is above 95%.
- No nuclease or RNase contamination.
- No residual host genomic DNA detected by PCR