

SARS-CoV-2 Inhibitor Screening Kit (RBD)

Catalog NO. : RK04194

version: 2.0

This package insert must be read in its entirety
before using this product

Introduction

This ELISA kit used for quantitative determination of SARS-CoV-2 Inhibitor Screening in serum and plasma. For research use only, and it' s highly recommended to read thoroughly of this manual before using the product.

Principle of the Assay

This assay is based on the competitive binding technique in which SARS-CoV-2 Inhibitor Screening present in a sample competes with ACE2 protein for a fixed amount of horseradish peroxidase (HRP)-labeled SARS-CoV-2 Inhibitor Screening. Following a wash to remove excess conjugate and unbound sample, a substrate solution is added to the wells to determine the bound enzyme activity. The color development is stopped and the absorbance is read at 450 nm. The intensity of the color is inversely proportional to the concentration of SARS-CoV-2 Inhibitor Screening in the sample.

Material Provided & Storage Conditions

Unopened kits can be stored at 2-8° C for 1 year, and opened products must be used within 1 month.

Part	Size	Cat. No.	Storage of opened/ reconstituted material
Antigen Coated Plate	8×12	RM41092	Put the unused slats back in the aluminum foil bag with the desiccant and reseal them. They can be stored at 2-8° C for 1 month.
Control Antibody (50×)	1 × 20 μ L	RM41093	It is not recommended to use again after redissolving.
Concentrated HRP- Conjugate Antigen (500×)	1 × 30 μ L	RM41094	Store at 2-8° c for 1 month *

Control/Sample Diluent (R1)	1 × 20mL	RM00023	Store at 2–8° c for 1 month *
HRP- Conjugate Antigen Diluent (R2)	1 × 12mL	RM00024	
Wash Buffer (20x)	1 × 30mL	RM00026	
TMB Substrate	1 × 12mL	RM00027	
Stop Solution	1 × 6mL	RM00028	
Plate Sealers	4 Strips		
Specification	1		

Other Supplies Required

1. Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 630 nm or 570 nm.
2. Pipettes and pipette tips.
3. Deionized or distilled water.
4. Squirt bottle, manifold dispenser, or automated microplate washer.
5. Incubator.
6. Test tubes for dilution of standards and samples.

Precautions

*** FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.**

1. Any variation in diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
2. Variations in sample collection, processing, and storage may cause sample value differences.
3. Reagents may be harmful, if ingested, rinse it with an excess amount of tap water.
4. Stop Solution contains strong acid. Wear eye, hand, and face protection.
5. Apart from the standard of kits, other components should not be refrigerated.
6. Please perform simple centrifugation to collect the liquid before use.
7. Do not mix or substitute reagents with those from other lots or other sources.
8. Adequate mixing is very important for good result. Use a mini-vortexer at the lowest frequency.
9. Mix the sample and all components in the kits adequately, and use clean plastic container to prepare all of the diluent.

10. Both the sample and standard should be assayed in duplicate, and the sequence of the reagents should be added consistently.
11. Reuse of dissolved standard is not recommended.
12. The kit should not be used beyond the expiration date on the kit label.
13. The kit should be away from light when it is stored or incubated.
14. To reduce the likelihood of blood-borne transmission of infectious agents, handle all serum, plasma and other biological fluids in accordance with NCCLS regulations.
15. To avoid cross contamination, please use disposable pipette tips.
16. Please prepare all the kit components according to the Specification. If the kits will be used several times, please seal the rest strips and preserve with desiccants. Do use up within 2 months.
17. This assay is designed to eliminate interference by other factors present in biological samples.
18. Until all factors have been tested in this assay, the possibility of interference cannot be excluded.
19. The 48T kit is also suitable for the specification.

Sample Collection & Storage

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Samples containing the correlated IgG as in this kit may interfere with this assay.

Serum : Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 xg. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Plasma : Collect plasma using EDTA or Heparin as an anticoagulant. Centrifuge for 15 minutes at $1000 \times g$ within 30 minutes of collection. Assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. (Note: Citrate plasma has not been validated for use in this assay.)

Note : It is suggested that all samples in a study be collected at the same time of the day. Avoid hemolytic and hyperlipidemia

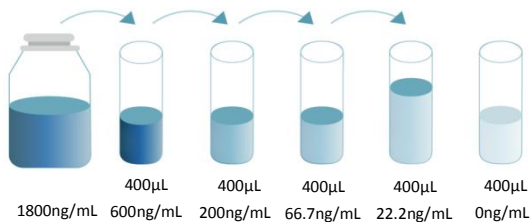
sample for Serum and Plasma.

Reagent Preparation

Bring all reagents to room temperature before use. If crystals have formed in the concentrate, Bring the reagent to room temperature and mix gently until the crystals have completely dissolved.

Control Antibody- Reconstitute the Control Antibody 50-fold dilution with Control/Sample Diluent(R1). This reconstitution produces a stock solution of 1800ng/mL. Mix the standard to ensure complete reconstitution and allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Use the 1800ng/mL standard stock to produce a dilution series (below) with Control/Sample Diluent(R1). Mix each tube thoroughly and change pipette tips between each transfer (recommended concentration for standard curve: 1800, 600, 200, 66.7, 22.2, 0ng/mL). Use diluted standards within 60 minutes of preparation.

Std 200 μ L 200 μ L 200 μ L 200 μ L



Concentrated HRP- Conjugate Antigen(500×) - Dilute 1:500 of Concentrated HRP- Conjugate Antigen (500x) with HRP- Conjugate Antigen Diluent(R2) before use, for example: Add 4 µL of Concentrated HRP- Conjugate Antigen (500x) to 1996 µL HRP- Conjugate Antigen Diluent(R2) to prepare 2000 µL Working HRP- Conjugate Antigen Buffer.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 1:20 with double distilled or deionized water before use, for example : Add 20 mL of Wash Buffer Concentrate to 380 mL of deionized or distilled water to prepare 400 mL of Wash Buffer.

Sample Preparation

Note – During the whole procedure, it is recommended that a face mask and gloves be used to protect kit reagents from contamination.

Sample – Different types of sample need appropriate dilutions. Serum (pretreated), plasma samples require a 5-fold dilution with Control/Sample Diluent (R1).

Assay Procedure

Bring all reagents and samples to room temperature before use. It is recommended that all controls, and samples be assayed in duplicate.

1. Prepare all reagents, working controls, and samples as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 50 μ L Control/sample Diluent (R1) in a blank well.
4. Add 50 μ L different concentration of Control and sample in other wells, record the plate layout of standards and sample assay.
5. Add 50 μ L Working HRP-Conjugate Antigen in each wells, cover with new adhesive strip provided.
6. Incubate for 1 hour at 37° C on a horizontal orbital microplate shaker (0.12" orbit) set at 500 \pm 50 rpm. During the incubation, turn on the microplate reader to warm up.
7. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with 350 μ L Wash Buffer using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at

each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.

8. Add 100 μ L TMB Substrate to each well. Incubate for 10–15 minutes at 37° C .Protect from light.
9. Add 50 μ L Stop Solution, determine the optical density of each well within 5 minutes, using a Microplate reader set to 450 nm. If wavelength correction is available, set to 570 nm or 630 nm. If wavelength correction is not available, subtract readings at 570 nm or 630 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

Assay Procedure Summary

Prepare the standard and reagents



Add 50ul of Control and test samples to each well



Add 50ul Working HRP-Conjugate Antigen



Incubate for 1 hour at 37° C, then wash 5 times



Add 100ul Substrate Solution



Incubate for 10-15 min at 37° C under dark condition



Add 50ul Stop Solution



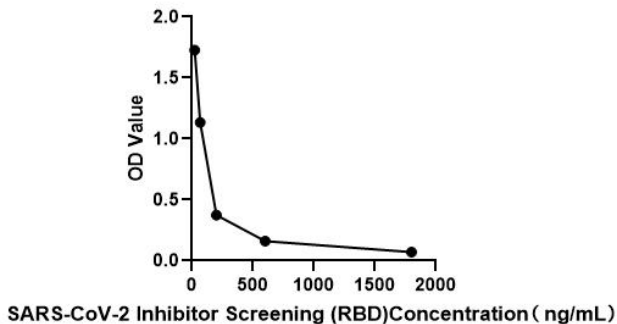
Detect the optical density within 5 minutes under 450nm.

Correction Wavelength set at 570nm or 630nm

Calculation of Results

1. Average the duplicate readings for each standard, control and sample, and subtract the average zero standard optical density (O.D.).
2. Create a standard curve by reducing the data using computer software capable of generating a four-parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the Y-axis against the concentration on the X-axis and draw a best fit curve through the points on a log/log graph. The data may be linearized by plotting the log of the SARS-CoV-2 Inhibitor Screening concentrations versus the log of the O.D. on a linear scale, and the best fit line can be determined by regression analysis.
3. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

Typical Data



The standard curves are provided for demonstration only. A standard curve should be generated for each set of SARS-CoV-2 Inhibitor Screening assayed.

Detection Range

22. 2-1800ng/mL

Sensitivity

The minimum detectable dose (MDD) of SARS-CoV-2 Inhibitor Screening typically less than 15.1ng/mL. The MDD was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

Specificity

This assay recognizes SARS-CoV-2 Inhibitor Screening .No significant cross-reactivity was observed.

Note:

Limited by current skills and knowledge, it is impossible for us to complete the cross-reactivity detection between SARS-CoV-2 Inhibitor Screening and all the analogues, therefore, cross reaction may still exist.

Precision

Intra-plate Precision

3 samples with low, middle and high level SARS-CoV-2 Inhibitor Screening were tested 20 times on one plate, respectively.

Intra-Assay: CV<10%

Inter-plate Precision

3 samples with low, middle and high level SARS-CoV-2 Inhibitor Screening were tested on 3 different plates, 20 replicates in each plate.

Inter-Assay: CV<15%

	Intra-Assay Precision			Inter-Assay Precision		
Sample	1	2	3	1	2	3
n	20	20	20	20	20	20
Mean (ng/mL)	570	930	1450	590	940	1480
Standard deviation	25.65	33.48	71.05	37.76	54.52	105.08
CV (%)	4.5	3.6	4.9	6.4	5.8	7.1

Trouble Shooting

Problem	Possible Cause	Solution
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High Background	Insufficient washing	Sufficiently wash plates as required. Ensure appropriate duration and number of washes. Ensure appropriate volume of wash buffer in each well.
	Incorrect incubation procedure	Check whether the duration and temperature of incubation are set up as required.
	Cross-contamination of samples and reagents	Be careful of the operations that could cause cross-contamination. Use fresh reagents and repeat the tests.
No signal or weak signal	Incorrect use of reagents	Check the concentration and dilution ratio of reagents. Make sure to use reagents in proper order.
	Incorrect use of microplate reader	Warm the reader up before use. Make sure to set up appropriate main wavelength and correction wavelength.
	Insufficient colour reaction time	Optimum duration of colour reaction should be limited to 15-25 minutes.
	Read too late after stopping the colour reaction	Read the plate in 5 minutes after stopping the reaction.
	Matrix effect of samples	Use positive control.

Too much signal	Contamination of TMB substrate	Check if TMB substrate solution turns blue. Use new TMB substrate solution.
	Plate sealers reused	Use a fresh new sealer in each step of experiments.
	Protein concentration in sample is too high	Do pre-test and dilute samples in optimum dilution ratio.
Poor Duplicates	Uneven addition of samples	Check the pipette. Periodically calibrate the pipette.
	Impurities and precipitates in samples	Centrifuge samples before use.
	Inadequate mixing of reagents	Mix all samples and reagents well before loading.

*For research purposes only. Not for therapeutic or diagnostic purposes.